

# Empirical methods in phonological research

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# Outline

General issues

Corpus techniques

A corpus example

A second corpus example

A third corpus example

Experimental techniques

An experiment example

A second experiment example

Conclusions

# Data collection in phonology

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## 1. Grammaticality judgments

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2. Fieldwork

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Most of these are less than optimal for phonological work. They are mostly written and typically **not** transcribed.

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- ▶ What does it cost? What can be gotten for free?

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  - ▶ Generic search tools, e.g. `grep`, `findstr`, etc.
  - ▶ Programming tools, e.g. Perl, Awk, etc.

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  - ▶ Phonology.

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- ▶ Is it appropriate for your goals?
- ▶ Do you have appropriate tools to find what you want?
- ▶ How do you **know** you've found something?

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- ▶ Let's make some commitment about corpus and tools.
  - ▶ Brown corpus for data
  - ▶ `findstr/grep` to do the counts
  - ▶ R for statistics
- ▶ These choices are for **pedagogical** purposes.

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- ▶ What does “more frequent than” mean?

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- ▶ We should then also throw out [g] because we want the same number of "categories".
- ▶ This is rather convenient because <g> maps to both [g] and [j].
- ▶ We've chosen sounds that map relatively directly to English spelling, but we still have to deal with some oddities, e.g. words like *know* [no].

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  - ▶ `findstr "[kn Kn mnMN]" bc.txt > mnwords.txt`

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  - ▶ Use MSWord to massage the `browncorpus.txt` file into something searchable. Use the global search-and-replace function to put each word on a separate line by replacing spaces with a return.
  - ▶ Use `findstr` in the DOS window to find lines that match the pattern, and pipe the results to a text file:
  - ▶ `findstr "^ [bdBD]" bc.txt > bdwords.txt`
  - ▶ `findstr "^kn ^Kn ^[mnMN]" bc.txt > mnwords.txt`
  - ▶ Open the resulting text file in Notepad, making sure the 'status line' is displayed, and move the mouse to the last line of the file.

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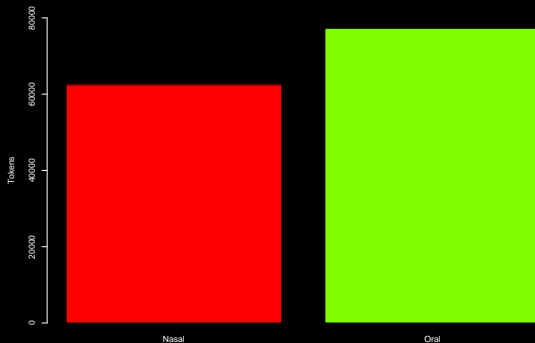
- ▶ Command-line interface
- ▶ Search expressions (regular expression syntax)
- ▶ Tool chains (piping and redirection)

# Results

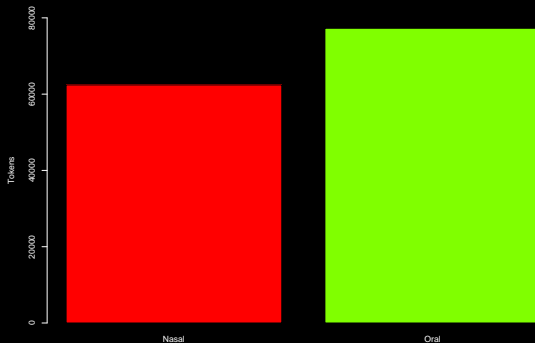
total number of words	1,026,595
words beginning with [mn]	62,415
words beginning with [bd]	77,262

# What does this look like?

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```
> barplot(c(62415,77262))
```

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Chi-squared test for given probabilities
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data: c(62415, 77262)
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X-squared = 1578.165, df = 1, p-value < 2.2e-16
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```
▶  $p$  is really small, much smaller than .05; our hypothesis is correct.
```

## Another corpus question

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- ▶ This can be tested more easily with a dictionary, rather than a text.
- ▶ For Unix geeks, there is another way to go: `grep -Eo pattern file | sort | uniq | wc`

# Newdic

## Newdic

- ▶ 20,000 words with various bits of information in tab-separated columns, e.g. transcription, stress, syllable count, spelling, frequency, part of speech.

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- ▶
 

x	_	S1	a	23178	(N IA VB PP)
ardvark	'	S2	aardvark	0	(N)
xb@k	_'	S2	aback	2	(AV)
@bxkxs	'__	S3	abacus	0	(N)
xb@ft	_'	S2	abaft	0	(AV PP)
@bxloni	'__	S4	abalone	0	(N)
xb@nd n	'__	S3	abandon	17	(VT N)
xbes	_'	S2	abase	0	(VT)
xb@S	_'	S2	abash	0	(VT)
xbet	_'	S2	abate	0	(VT VI)

# How to count

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- ▶ Easier because each word is on a separate line.
- ▶ We need to count only those items where the relevant letter is at the beginning of the line.

# Details

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- ▶ Mac or Unix

- ▶ `grep -Ec '^[mn]' newdic.txt`
- ▶ `grep -Ec '^[bd]' newdic.txt`

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- ▶ `grep -Ec '^[mn]' newdic.txt`
- ▶ `grep -Ec '^[bd]' newdic.txt`

## ▶ Windows

- ▶ `findstr "^[bd]" newdic.txt > bdwords.txt`
- ▶ `findstr "^[mn]" newdic.txt > mnwords.txt`
- ▶ Open resulting files with Notepad

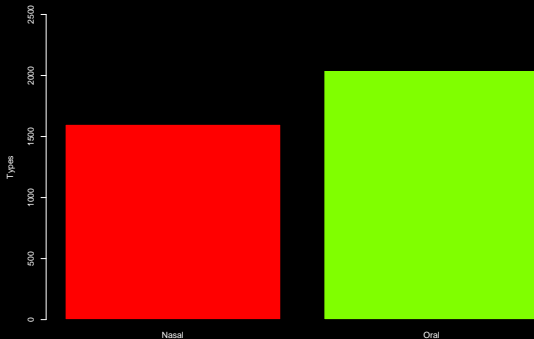
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total number of words	19,528
words beginning with [bd]	2,041
words beginning with [mn]	1,600

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> barplot(c(1600,2041))
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# Statistics

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Chi-squared test for given probabilities
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```
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X-squared = 53.4142, df = 1, p-value = 2.701e-13
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```

- ▶ There is a significant difference in the number of distinct words that begin with either a nasal or a voiced stop.

# A third corpus example

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- ▶ Is there a difference in the number of types (distinct words) when we factor in the number of syllables in the word?
- ▶ Again, we can do this better with a dictionary.
- ▶ The `newdic` dictionary is convenient as the third column indicates the number of syllables: S1, S2, S3, etc.

# What to count

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- ▶ For Unix or Mac: `grep -Ec '^[mn].*S1' newdic.txt`
- ▶ For Windows: `findstr "^[mn].*S1" newdic.txt > mnwords.txt`, and open with Notepad.

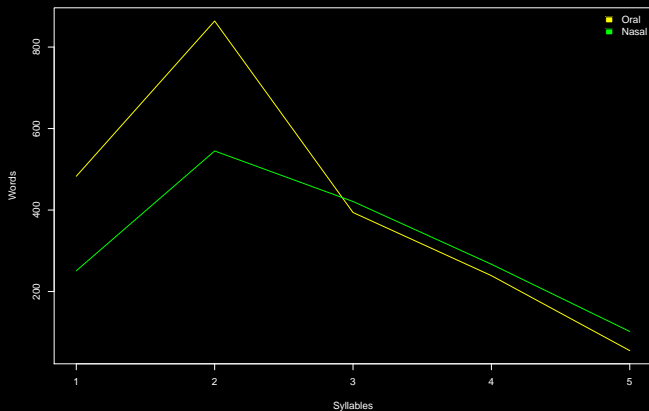
# Results

Syllables	Nasal	Voiced
1	251	483
2	545	864
3	421	394
4	267	239
5	102	55

## Results in a form R understands

Nasal	Voiced
251	483
545	864
421	394
267	239
102	55

# Results graphed



## Reading in and plotting the data

```
> sno <- read.table('sylnasor.txt',header=T)

> plot(sno$Voiced,type='l',col='yellow',
       xlab='Syllables',ylab='Words')

> lines(sno$Nasal,col='green')

> legend('topright',legend=c('Oral','Nasal'),
       fill=c('yellow','green'))
```

# Stats

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```

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```
X-squared = 1.5494, df = 1, p-value = 0.2132
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X-squared = 0.8945, df = 1, p-value = 0.3443
```

▶ **Four syllables:**

```
> chisq.test(sno[4,])
X-squared = 1.5494, df = 1, p-value = 0.2132
```

▶ **Five syllables:**

```
> chisq.test(sno[5,])
X-squared = 14.0701, df = 1, p-value = 0.0001761
```

# What does it all mean?

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- ▶ The latter generalization holds for one- and two-syllable words, but does not hold for three- and four-syllable words. The pattern reverses itself with five-syllable words.
- ▶ Fame and fortune is yours if you can figure out why. . .

# Doing experiments

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- ▶ Kinds of experiments

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- ▶ Materials

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- ▶ Kinds of experiments
- ▶ Materials
- ▶ Human subjects

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- ▶ Collect judgments of grammaticality
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Are these relevant if one accepts a distinction between “competence” and “performance”?

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- ▶ How many items in each group? Depends on the size of the effect, the amount of statistical noise expected, what statistics one will run, etc.
- ▶ Sometimes one has to include irrelevant items: **distractors**. This is so subjects don't become aware of the contrast being investigated.

# Human subjects

In the US, there is a **huge** hurdle at this point: IRB approval.

## A hypothesis

H<sub>x</sub>: people prefer words that begin with voiced stops to words that begin with nasals.

# Experimental design

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- ▶ Visual presentation, in English orthography
- ▶ Paired items
- ▶ Include distractors

# Experimental items

	oral	nasal
labial	barna, bimp, bulrow	marna, mimp, mulrow
coronal	dift, dact, dilsey	nift, nact, nilsey

distractors: keff, farn, skelsat, starno, also, plent

# Free software

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- ▶ 3 × 5 cards

# How I did it

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The version of the webpage that you can download is an **offline** version. It collects responses, but the experimenter must explicitly save them.

# What we have to do

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- ▶ Conceptually, we have to:
- ▶ Practically, we have to:

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- ▶ Conceptually, we have to:
  - ▶ figure out what differences may have shown up; and
  - ▶ figure out if they are **significant**.
- ▶ Practically, we have to:
  - ▶ get the data into R;
  - ▶ calculate and plot the differences; and
  - ▶ figure out if they are **significant**.

# Massaging the results

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- ▶ A form that we can get into R.

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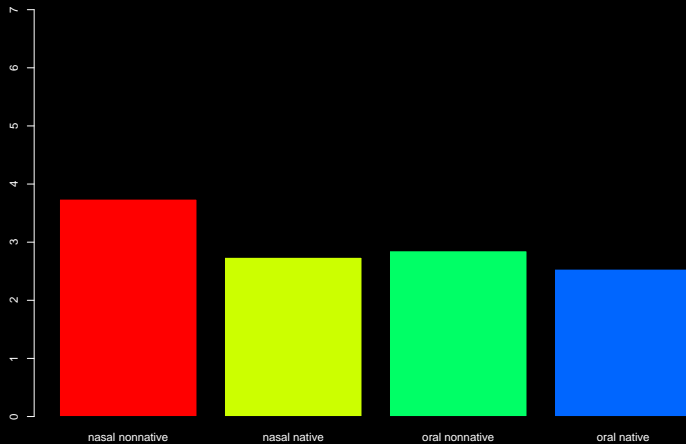
- ▶
 

subject	native	place	nasal	item	response
s1	no	lab	nas	marna	4
s1	no	cor	nas	nact	1
s1	no	cor	nas	nilsey	2
s1	no	lab	oral	dact	1
s1	no	lab	oral	bimp	1
s1	no	cor	nas	nift	1
s1	no	lab	oral	barna	4
s1	no	lab	oral	dift	1
s1	no	lab	nas	mullow	1

## Getting the data into R

```
> taiexp <- read.table('results.txt',header=T)
> summary(taiexp)
...
> attach(taiexp)
> tapply(response,list(native,nasal),mean)
      nas      oral
no  3.740741 2.851852
yes 2.739130 2.536232
```

# Plotting the means



## Creating that chart

```
> attach(taiexp)
> the.means <- tapply(response,list(native,nasal),mean)
> barplot(as.vector(the.means), ylim=c(0,7),col=rainbow(5),
names.arg=c('nasal nonnative', 'nasal native',
'oral nonnative','oral native'))
```

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- ▶ Here we must do an “Analysis of Variance” (ANOVA).
- ▶ **Warning:** this is probably the wrong statistic to perform, but it’s what the literature uses, so it’s what we have to use.

# ANOVA

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- ▶ Determine all the variables and all the possible interactions of variables.

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- ▶ Calculate the statistical noise (variance).

# ANOVA

- ▶ Determine all the variables and all the possible interactions of variables.
- ▶ Factor out the random variables: subjects and items.
- ▶ Calculate the difference between conditions.
- ▶ Calculate the statistical noise (variance).
- ▶ Is the difference greater than the noise ( $F$  ratio)?

## Factoring out subject noise

```
> summary(aov(response ~ native * nasal +
Error(subject/(native * nasal))))
Error: subject
          Df Sum Sq Mean Sq F value Pr(>F)
native    1  39.18   39.18   5.1995 0.02664 *
Residuals 53 399.39    7.54
Error: subject:nasal
          Df Sum Sq Mean Sq F value Pr(>F)
nasal     1 16.388  16.388 10.9964 0.001653 **
native:nasal 1 10.627  10.627  7.1306 0.010040 *
Residuals  53 78.986    1.490
```

## Factoring out item noise

```
> summary(aov(response ~ native * nasal +
Error(item/(native * nasal))))
Error: item
      Df Sum Sq Mean Sq F value Pr(>F)
nasal  1  16.39   16.39  0.4965 0.4971
Residuals 10 330.08   33.01
Error: item:native
      Df Sum Sq Mean Sq F value  Pr(>F)
native  1 39.182  39.182 15.8293 0.002607 **
native:nasal  1 10.627  10.627  4.2931 0.065066 .
Residuals    10 24.753   2.475
```

## A second experiment

Hx: people prefer words that begin with voiced stops to words that begin with nasals. This will show up with yes–no judgments.

# How do we do this?

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- ▶ Web-based experiment parallel to the last one, but changing the response categories to yes–no.

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- ▶ Web-based experiment parallel to the last one, but changing the response categories to yes–no.
- ▶ Do we expect a different outcome?
- ▶ 50 subjects participated.

# Categorical response

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## Categorical response

- ▶ Yes–no judgments are a **categorical** response, not a quantitative response.
- ▶ We need to convert our data into a quantitative measure so we can do an ANOVA.
- ▶ Aggregate across places of articulation and multiple items within each condition.
- ▶ We can then do a by-subjects ANOVA.
- ▶ But we **cannot** do a by-items ANOVA.

## Results massaged

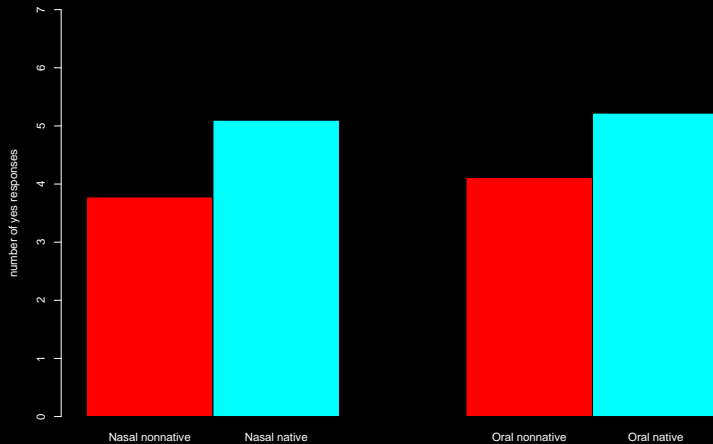
subject	native	nasal	response
s1	yes	nas	6
s1	yes	oral	6
s10	yes	nas	6
s10	yes	oral	6
s11	yes	nas	4
s11	yes	oral	4
s12	yes	nas	6
s12	yes	oral	6

# Means

	nas	oral
no	3.777778	4.111111
yes	5.097561	5.219512

```
> te2 <- read.table('res2a.txt',header=T)
> attach(te2)
> tapply(response,list(native,nasal),mean)
```

## Plotting the means



## How we did that

```
> attach(te2)
> barplot(tapply(response,list(native,nasal),mean),
beside=T,ylim=c(0,7),col=rainbow(2),
names.arg=c('Nasal nonnative','Nasal native',
'Oral nonnative','Oral native'),
ylab='number of yes responses')
```

## Stats

```
> summary(aov(response ~ native * nasal +
Error(subject/(native * nasal))))
```

```
Error: subject
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
native	1	21.757	21.757	7.6854	0.0079 **
Residuals	48	135.883	2.831		

```
...
```

```
Error: subject:nasal
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
nasal	1	0.6400	0.6400	1.6004	0.2120
native:nasal	1	0.1649	0.1649	0.4123	0.5239
Residuals	48	19.1951	0.3999		

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- ▶ There are very important practical realities to doing any kind of experimental work, e.g. selecting a corpus, selecting the right tools, designing an experiment.
- ▶ You need **some** knowledge of statistics.
- ▶ We've **barely** scratched the surface here.